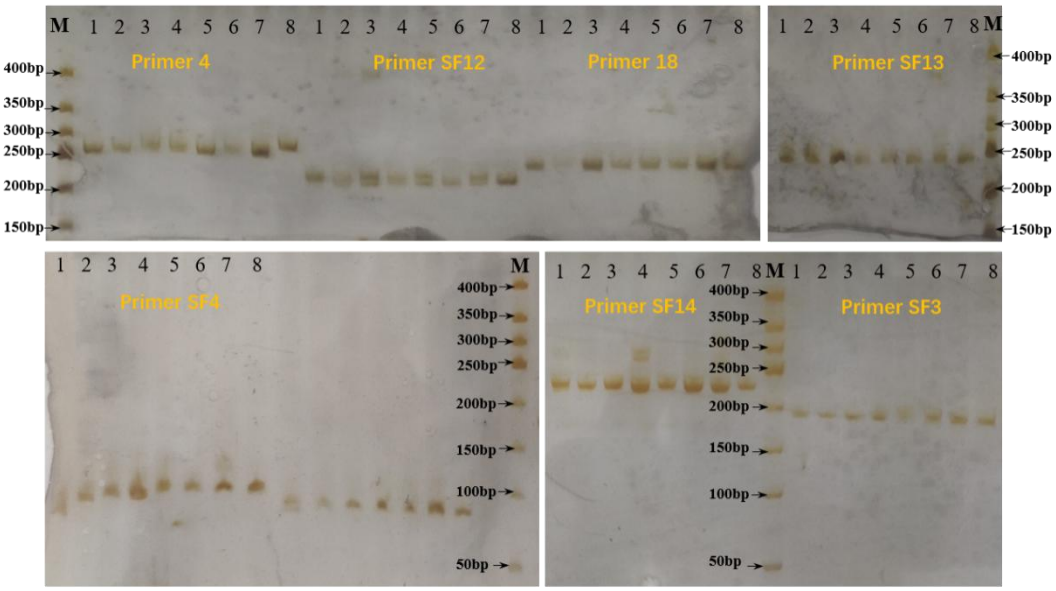


Figure 5 Amplification and detection of primers 4, 18, SF3, SF14, SF4, SF12 and SF13
Note: 1~8 The number of strains; M: 50bp DNA ladder

Table 6 Information of microsatellite primers used in this study

| Locus name | Dye | Primer sequence (5′-3′) | Tm (℃) | Allele size (bp) | Repeated motif |
|---|---|---|---|---|---|
| SF12 | 5`6-FAM | F:AAACGAATGAGGCTGAATGG | 58 | 150-200 | $(AT)_6$ |
| | | R:GGATGCACGCAATTTTCTTT | | | |
| SF3 | 5`6-FAM | F:GAGAGCTCTGCTGCCATCTT | 58 | 150 | $(TC)_6$ |
| | | R:ATAACGTTCCCTGGCATCTG | | | |
| SF4 | 5`6-FAM | F:ATAAAAAGTCCCCGGAGCAT | 58 | 100-150 | $(AG)_9$ |
| | | R:GCCAGTGAAATTGAGGTTGC | | | |
| 18 | 5`6-FAM | F:ACATTGATTTGCATTGGGGT | 58 | 200-250 | $(CA)_6$ |
| | | R:AGAGCACATTCCGGTACCAC | | | |
| SF13 | 5`HEX | F:ACGGCCTTTGTTTTCTCTCA | 58 | 250-300 | $(GT)_7$ |
| | | R:AAACCGCCAACACAGGTAAT | | | |
| SF14 | 5`HEX | F:CTTCGTCCCCGATACAAGAG | 58 | 200-300 | $(CAG)_6$ |
| | | R:CATCATGCCCGATATCATCA | | | |
| 4 | 5`HEX | F:AGTGAGAGCACCTGCTGGAT | 58 | 300 | $(TTC)_5/(GGGTAAA)_3$ |
| | | R:AGCAGTGGGCTTTACCCTTT | | | |

Table 7 The PCR reaction system of the microsatellite markers

| Component | Volume (μL) |
|---|---|
| (Vazyme)2×Taq Master mix | 12.5 |
| Forward primer | 1.5 |
| Reverse primer | 1.5 |
| $ddH_2O$ | 6 |
| DNA template | 1.5 |
| Total | 20 |

The PCR reaction program was as follows: 94 °C for 3 min; 30 cycles of 94 °C for 30 s, 58 °C for 30 s, and 72 °C for 1 min; followed by a final extension at 72 °C for 5 min, and then held at 4 °C. After completion of the PCR reactions, the products were examined by electrophoresis on 3% agarose gels. Qualified PCR amplification products were sent to an automated sequencer (Applied Biosystems) for allele genotyping. GeneMarker software was used to read allele sizes, and genotyping results were obtained for 100 *Platycladus orientalis* individuals.